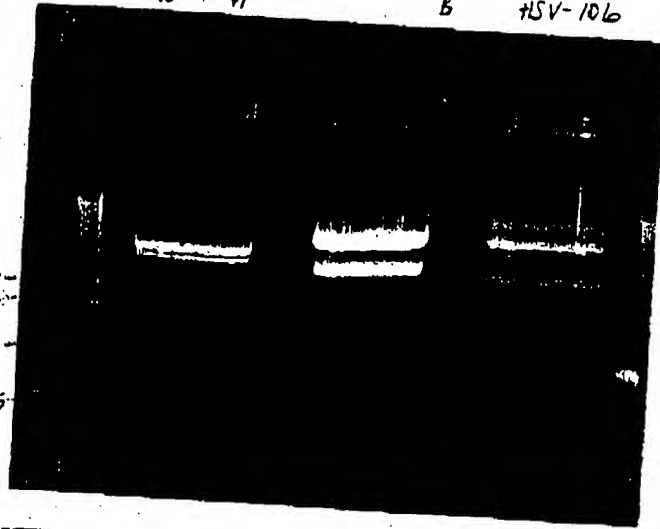


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15.90 A

B

HSV-106



15.90 A: BspHI never cut
even w/ o/n of extra
enzyme = β -gal = Xba
took upper band

15.90 B: XbaI complete on
took upper band

also 15.91
took band at ~ 2.2 kb

did ~~Agar~~ Glacommilic Na2
control. Eluted w/ 20 μ l x
H₂O and froze o/n

15.90a
15.90b
HSV106



End of each sample for concentration re

$$1. \quad 15.90 A = 100 \mu\text{g} / 5 \mu\text{l} = 20 \mu\text{g} / \mu\text{l} = \beta\text{-gal}$$

want 34.2 ng for ligation = 1.5 μ l

$$2. \quad 15.90 B = \text{XbaI} / \text{HindIII PstE}$$

$$= 0.6 \mu\text{g} / 5 \mu\text{l} = 120 \text{ ng} / \mu\text{l}$$

want 45 ng = 0.5 μ l

$$3. \quad 15.91 = \text{HSV-tk gene}$$

$$= 10 \text{ ng} / \mu\text{l}$$

want 20 ng = 2 μ l

Vectra 15.81 A use 2 μ l of 1:5 diln

Vectra 15.81 B use 2 μ l of 1:5 diln

pCDNA p16.7L want 1.8 ng use 2 μ l of 1:100



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- ligate a) vector p15.812 p-gal int. PS as Xba1
 insert p15.90a
- b) vector p15.812 p50 to PS as Xba1 (blunt) / HindIII
 insert p15.90b into HindIII / HindIII PS 1cs
- c) ~~vector~~
~~insert~~ p15.91 HSV-FR int pEDNA3 as BglII / EcoRI
~~insert~~ p16.71 into BamHI / EcoRI

GenI set-up the ligation last evening @ 14°C o/n
 this morning - run ligation control gel
 - plate in 50 µg/ml Amp^r lanes 1, 2, 5-16

yesterday I also re-transformed the four additional clones
 from Kate Microbia, Toronto, ON. They failed to
 grow again - not much more than this morning. I will
 try one last time today using the ligation experiment
 as a control.

Also want to characterize the following plasmids from Kate
 that came out of Cell yesterday

pΔE15p13 (lower band from Cell)

use 2 µl / digest

pXCI (lower band Cell)

use 2 µl / digest

pBHG 10 or 11

use 5 µl / digest

← this doesn't look as
 good as others

note: pDidi 5.0a doesn't look too good at all
 & needs to be digested

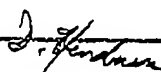
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